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## Mutation in Virus and Genetic Alteration in Viruses - A Review

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**Abstract:** The remarkable capacity of some viruses to adjust and adapt to a new host and environment is highly dependent on the capacity to generate de Novo diversity in a short period. Rates of spontaneous mutation shift abundantly among viruses. RNA viruses change quicker than DNA viruses, single-stranded virus transforms quicker than double standard infection and genome size seems to connect adversely with transformation rate. Viral change rates are equalised at various stages, like polymerase fidelity, Sequence content, template secondary structure, cellular microenvironment, application mechanism, proofreading and post replication repair. The mutation rate of an organism is defined as the probability where a change in genetic information is passed to the next generation. In viruses, a generation is often defined as a cell infection cycle, which includes an attachment to the cell surface, entry, gene expression, replication, encapsidation, and release of infectious particles. Also, enormous quantities of changes can be presented by some virus-encoded diversity generating elements, as well as by host-encoded cystine/ adenosine deaminase. Our recent knowledge on viral change rates explains that viral hereditary variety is determined by multiple viruses and host interaction and the viral mutation rates can evolve in response to specific selective pressures.

**Keywords:** Virus; Mutation rates; Radiation induced DNA damage; Cross species adaptation in viruses; Food preservatives; Recombination

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### INTRODUCTION

The mutation rate of an organism is defined as the probability where a change in hereditary information is passed to the next generation. In viruses, a generation is frequently characterised by a cell infection cycle, which includes an attachment to a cell surface, entry, gene expression, replication, encapsidation and arrival of infection particle (Sanjuán, 2016). Changes are not confined to replication since they alter the material or unconstrained nuclei causing harm (Duffy, Shackelton and Holmes, 2008). The change rate ought not to be mistaken for the recurrence at which transformation is found in a given viral population (Perelson, 2018). The last is a proportion of hereditary variety that relies upon various procedures, for example, characteristic choices, arbitrary hereditary float, recombination etc. Viral mutation rates are not merely caused by polymerase crosses, but also by the ability of a virus to correct DNA mismatches by proofreading and post replicative repair (Schotsaert and García-Sastre, 2014). Mutation rates are modulated by additional factors, including proteins involving in the application other than the polymerase the node of replication, template sequence and structure (Cao *et al.*, 2016). Our team has rich experience in research and we have collaborated with numerous authors over various topics in the past decade (Deogade, Gupta and Ariga, 2018; Ezhilarasan, 2018; Ezhilarasan, Sokal and Najimi, 2018; Jeevanandan and Govindaraju, 2018; J *et al.*, 2018; Menon *et al.*, 2018; Prabakar *et al.*, 2018; Rajeshkumar *et al.*, 2018, 2019; Vishnu Prasad *et al.*, 2018; Wahab *et al.*, 2018; Dua *et al.*, 2019; Duraisamy *et al.*, 2019; Ezhilarasan, Apoorva and Ashok Vardhan, 2019; Gheena and Ezhilarasan, 2019; Malli Sureshbabu *et al.*, 2019; Mehta *et al.*, 2019; Panchal, Jeevanandan and Subramanian, 2019; Rajendran *et al.*, 2019; Ramakrishnan, Dhanalakshmi and Subramanian, 2019; Sharma *et al.*, 2019; Varghese, Ramesh and Veeraiyan, 2019; Gomathi *et al.*, 2020; Samuel, Acharya and Rao, 2020)

## MATERIALS AND METHODS

A systematic review of scientific literature was done in preparation of the manuscript. The systems and databases searched from relevant articles were from PUBMED, Google scholar and WHO online articles. Databases of intended journals were searched for keywords such as COVID-19, preventive measures, safety, transmission, treatment etc. The exclusion criteria were case reports review and studies in other languages.

## DISCUSSION

The transformation/ Mutation rate is a basic parameter for understanding viral development and has a significant functional implication. For example, the range of the mutation rate of HIV-1 demonstrated that any single change giving medication obstruction ought to happen and that conventional treatment with various medications was subsequently necessary (Anderson, Daifuku and Loeb, 2004). Additionally, in principle information with high transformation rates could be battled by the organisation of mutagens (Bull, Sanjuán and Wilke, 2007). The technique, called Lethal mutagenesis has demonstrated success in cell culture or animal models against RNA viruses, including enterovirus. The viral mutation rate also plays a role in the preparation of possible vaccination (Domingo *et al.*, 2006). Finally, at both the epidemiology and evolutionary levels, the mutation rate is one of the factors that can determine the risk of emerging infectious disease pathogens crossing the species barrier (Crotty, Cameron and Andino, 2001). Single slight changes in extended-spectrum can also determine whether or not some virus infections are cleared by the host immune system and can produce dramatic differences in virus fitness and virulence (Holland *et al.*, 1990). However, our knowledge about viral mutation rate is somewhat incomplete, due to the difficulty in measuring inheritance that occurs randomly and also due to several sources of bias, inaccuracies and terminological confusion (Graci *et al.*, 2007; Ashwin and Muralidharan, 2015). Orthodontic appliances are placed in the patient's oral cavity for a longer period of time. It will affect the oral hygiene of the patient. Oral prophylaxis is very important to prevent complication during the treatment. Physical removal of debris is difficult in such case mouth rinses are widely used to kill the bacteria (Website, no date a, Website, no date b).

### Mutation in viruses

The mutation rate of an organism is defined as the probability that a change in genetic information is passed to the next generation. In viruses, a generation is often defined as a cell infection cycle, which includes an attachment to the cell surface, entry, gene expression, replication, encapsidation, and release of infectious particles. Mutations are not restricted to replication since they can also result from the editing of the genetic material or spontaneous nucleic acid damage. The mutation rate should not be confused with the frequency of mutations which occurs in a given viral population. The latter is a measure of genetic variation that depends on several other processes such as natural selection, random genetic drift, recombination, and so on. Higher mutation rates lead to higher genetic diversity but, except in special cases, it is not possible to interfere with mutation rates directly from observed population mutation frequencies (Belshaw, Sanjuán and Pybus, 2011). *Acinetobacter baumannii* associated with nosocomial infections are among the top six drug resistant microbes. Extensive use of the  $\beta$ -lactam group of antibiotics has resulted in the emergence of drug resistance and has raised a major clinical crisis. Among the newer  $\beta$ -lactamases, extended spectrum  $\beta$ -lactamases (ESBLs) have emerged as a major cause of resistance against cephalosporins (Website, no date c, Website, no date d, Website, no date e). Although genetic diversity depends on multiple factors, the mutation rate is of particular interest because it constitutes the ultimate source of genetic variation. Point mutation, for the most part, occurs during DNA replication. DNA replication happens when one twofold abandoned DNA atom makes two single strands of DNA, every one of which is a layout for the making of the corresponding strand. A solitary point transformation can change the entire DNA arrangement. Transforming one purine or pyrimidine may change the amino corrosive that the nucleotides code for.

Point changes may emerge from unconstrained transformations that happen during DNA replication. The pace of transformation might be expanded by mutagens. Mutagens can be physical, for example, radiation from UV beams, X-beams or extraordinary warmth, or substance (atoms that lose base matches or disturb the helical state of DNA). Mutagens related to malignant growths are regularly concentrated to find out about the disease and its counteraction (Garcin, Itoh and Kolakofsky, 1997; Geetha, Thangavelu and Others, 2019). There are different ways to guide changes to happen. To begin with, bright (UV) light and higher-recurrence light are fit for ionizing electrons, which thus can influence DNA. Receptive oxygen atoms with free radicals, which are a side-effect of cell digestion, can likewise be unsafe to DNA. These reactants can prompt both single-abandoned DNA breaks and twofold abandoned DNA breaks. Third, bonds in DNA, in the end, debase, which makes another issue to keep the trustworthiness of DNA an exclusive requirement. There can likewise be replication blunders that lead to replacement, inclusion or ensure transformations (Peace-Brewer *et al.*, 1996).

### **Transfer of genetic elements from virus**

The genomes of huge eukaryotic twofold abandoned DNA infections contain high extents of cell qualities because of host-to-infection level exchanges (HT). For instance, at any rate, 10% of goliath infection qualities and up to 30% of herpesvirus qualities likely started from eukaryote or prokaryote genomes. A portion of these qualities has been appeared to go about as key factors in the aetiology of viral changes, because cell quality substance can be very extraordinary between firmly related infections as well as very comparable between indirectly related infections viral co-choice of host qualities seems, by all accounts, to fairly visit during infection development (Gilbert *et al.*, 2016). The cell qualities that have so far been recognized in viral genomes result from generally antiquated host-to-infection HT occasions. In concurrence with this speculation, a considerable lot of these qualities are thought to assume a job in upsetting hosts hostile to viral barriers, accordingly encouraging viral replication (Website, no date f). An end product of this situation is that numerous viral-borne have qualities coming about because of host-to-infection HT ought to be found at different frequencies in viral populaces. Many HT cases have been portrayed in eukaryotes. A large number of these exchanges have produced transformative oddities and permitted accepting creatures to adjust to new conditions. Even the exchange of DNA is in this manner progressively refreshing as a significant developmental power moulding eukaryotic genomes. Be that as it may, the components and the potential vectors associated with HT of DNA between eukaryotes remain ineffectively known, particularly in multicellular eukaryotes (Hughes and Friedman, 2005). Infections have been proposed as competitor vectors encouraging HT between eukaryotes since they are transmitted evenly (and now and again vertically) and they imitate inside host cells.

### **Radiation-induced DNA damage**

Ionising radiation induces genomic instability, which is transmitted over numerous ages after irradiation through the progeny of surviving cells. Induced genomic instability is shown as the symptom of delayed effects; delayed death or lethal mutation, chromosomal instability and mutagenesis. Since induced genomic instability causes gene mutation and rearrangement of chromosomes, it has been thought to play an important role in radiation-induced carcinogenesis (Adelman *et al.*, 1987). Radiation-induced genomic instability, shows its effect for prolonged periods, causing initial DNA damage in the surviving cells is memorised (Barcellos-Hoff and Brooks, 2001). In general, the frequency of a given mutation increases in proportion to the dose of radiation in the low to intermediate dose range (Chang and Little, 1991). At higher doses, however, frequency of mutations induced by a given dose may be dependent on the rate at which the doses accumulated, tending to be lower if the dose is accumulated over a long period (Cremer *et al.*, 2001). The capacity of radiation to increase the frequency of mutation is often expressed in terms of the mutation rate doubling dose, which is the dose that induces a large and additional rate of mutation as that which occurs spontaneously in each generation (Hill, 1999). The more sensitive the genes are to radiation, the lower is the doubling (Khanna *et al.*, 2001). Taking into account the way the radiation is currently accepted to play a role in mutagenic or cancer-causing action, any procedure involving radiation exposure is considered to impact some degree of risk (Liang *et al.*, 2002). At the same time, radiation-induced high risks associated with many activities are negligibly small in comparison with other ways commonly encountered in daily life (Clutton *et al.*, 1996). Systematic efforts are made to avoid unnecessary exposure to ionising radiation in medicine, science and industry (Podolin *et al.*, 2002).

### **DNA damage due to food preservatives**

The use of food additives has increased enormously in modern food technology but they have adverse effects on human health. Food additives are used widely for various purposes, including preservation, colouring and sweetening. They are added to stop or delay multi-nutritional losses due to microbiological, enzymatic or chemical changes of foods and to prolong the shelf life and quality of foods (Yılmaz, Ünal and Yüzbaşıoğlu, 2009). Recently food additives have attracted attention as a potential cause of various human diseases they might be among the factors responsible for the outbreak of cancer, hepatic and nephritic failures among mutagenic potential. (Tanaka, 2007; Pandir, 2016). The modern technologies in the food industry have resulted in the use of a variety of food additives alone in combination. There are many studies conducted to prove the effect of food preservatives in DNA and many studies have proved that there is DNA damage due to various food additives. Many studies have been conducted to see if there are any mutations taking place due to preservatives. Several studies suggested that certain types of azo dyes, including SY (sunset yellow), exhibit mutagenic activity. (Yılmaz *et al.*, 2008) The toxicity and carcinogenicity of SY in mammalian systems may result either via interactions of intact molecules with Cytosolic receptors or via the formation of free radicals and arylamines azo reduction. Benzoic acid is commonly used as an antimicrobial agent in many food products. Certain researchers have proved that there is a significant decrease in life periods and increases somatic mutation and recombination. It is shown that Benzoic acid is a weak genotoxic agent, especial at a lower concentration, in human lymphocytes (Biswas and Khuda-Bukhsh, 2005; Zengin *et al.*, 2011)

### **Cross-species adaptation in virus**

Model living organisms have been fundamentally significant in biochemical research, and are especially helpful for examining biological processes and pathways that operate by comparable guidelines across different species (Fadel and Poeschla, 2011). The dynamic interplay between host and virus in nature is hard to relate in research facility-based examinations that utilise a single viral clone infecting and isogenic host population (Planelles, 2012). First, in the process of virus-host switching, a virus of a species involves the ability to infect and spread in a second species. In this procedure, hereditary contracts between the species, not hereditary similitudes, are what direct the developmental adjustments required by the infections (Malim and Bieniasz, 2012). Second, infections and the host qualities that encode safeguards against them are known to be outstanding for their hereditary assorted variety. In this way, the after-effects of trials utilising clonal host and clonal infection in the research facility may not generally uncover the range of concernable host infection cooperation exists in nature (Meyerson and Sawyer, 2011). Third, in studies of viruses infecting their natural host species, including cell lines derived from those species, host defence mechanisms can be masked because the viruses have already evolved to evade them. In all of these interactions, experiments conducted in non-host species can be highly informative (Sayah *et al.*, 2004). Here we consider both the qualities and confinement of approaches including disease of heterologous creatures, heterozygotes cell lines and even cell lines contrasting just by the outflow of single qualities from heterologous species (Stremlau *et al.*, 2005). Impressive advancement has been made in distinguishing the numerous components that control or impact infection have exchanged. While it is as yet impractical to recognize which among the thousands of infections in wild or residential creatures will rise in people or precisely where and when the following developing zoonotic infections will start, contemplates point to the normal path (Malim and Bieniasz, 2012) ways and recommend preventive techniques. With better data about the birthplaces of new infections, it might be conceivable to recognize and control possible infections. Traditional disease control methodology, (for example, wellbeing checking and isolate) can considerably lessen contact among supply and beneficiary hosts, forestalling flare-ups or ending them after host move however while they are as yet constrained in size (Jefferson *et al.*, 2009). For arboviruses, vector control can constrain the transmission of infections from their repositories to a new host. There is questionable proof that general well-being measures under-taken in 1918 were compelling in controlling the flu of that year (Bootsma, 2007; Hatchett, Mecher and Lipsitch, 2007). Different methodologies include diminishing anthropogenic change in rising irresistible problem areas, just as the more costly and morally testing approach of separating repository creatures or the immunization of those creatures. Inoculation has been utilized effectively for standard trial control of rabies in the United States and Europe (by immunizing raccoons or foxes) and for control of wild pooch rabies in Kenya and Tanzania (by immunizing residential canines). New quickly spreading infections can get difficult to control once they cross the limit of a specific number of contaminations or potentially pace of transmission, for instance, after spreading in people into urban populaces, where isolation, as well as treatment, gets unfeasible (Bauch *et al.*, 2005; Paramasivam, Vijayashree Priyadharsini and Raghunandhakumar, 2020). Consequently, coordinated vital arranging is basic for the quick reactions required to defy new infections right on time after development. Such arrangements must be to some degree conventional because we come up short on the capacity to anticipate which infection will arise or what it's pathogenic. National and universal arrangements are additionally basic, including the saddling of logical and analytic advancements and building up techniques for rap-inertly imparting data about flare-ups and for co-ordinating control measures. Methodologies ought to incorporate improved reconnaissance focused to districts of high probability for the rise, improved recognition of pathogens in repositories or ahead of schedule in outbreaks, extensively based research to explain the significant advances that favour rise, and changed types of traditional or other control measures. Human sickness reconnaissance unmistakably should be related to improved longitudinal veterinary and wild-creature contamination observation (Martina *et al.*, 2003; Fouchier *et al.*, 2005). Antibody systems could be utilized in some control programs, however, the slow pace of improvement and endorsement of human immunizations is too low to even think about allowing control of most recently developing infection illnesses. Existing antibodies can be utilized to control the rise of known infections when adequate lead time is accessible, as might veterinary immunizations which can be grown generally rapidly and used to battle episodes, alongside the separating or quarantine measures that are currently regularly utilized. Better than ever antibody advances incorporate molecularly cloned constricted infections that can be quickly changed into the suitable enemy of genic structures with adequate viability and a degree of hazard low enough for use despite certain flare-ups. Antiviral medications might be utilized where accessible, even though cost, calculated issues, and symptoms may make those increasingly hard to use in a huge scope flare-up, and they would almost certainly work just with regards to other control measures (Fraser *et al.*, 2004; Ferguson *et al.*, 2005). The rise of new popular infections by creature-to-human host exchanging has been, and will probably keep on being, a significant wellspring of new human irresistible illnesses. A superior comprehension of the numerous intricate factors that underlie such rises is of most extreme significance to general wellbeing

### Recombination in Virus

Recombination is a prevalent process generating diversity in most viruses. It joins variations that emerge autonomously inside a simple particle, making new open doors for infection to adapt to new environments and hosts (Arenas, 2012; Girija, Priyadharsini and Paramasivam, 2019). Consequently, the study of viral recombination has attracted the interest of clinicians, epidemiologists, molecular biologists and evolutionary biologists. Viruses undergo genetic change by several mechanisms, including point mutation and recombination (Arenas and Posada, 2010; Priyadharsini *et al.*, 2018). In general RNA viruses have smaller genomes than DNA viruses some may even have DNA and RNA at different stages in their life cycle. The reason for this inverse relationship between genome size and mutation rate is arguably the incapability of large RNA viruses to replicate without generating lethal mutation. In contrast, DNA viruses generally have a larger genome of the higher fidelity of their replication enzymes (Awadalla, 2003). Recombination occurs when at least two viral genomes infect the same host cell and exchange genetic segments. Recombination is a widespread phenomenon in viruses and can have a major impact on their evolution (Baltimore, 1971). Recombination has been associated with the expansion of viral host ranges, the emergence of new viruses, the alteration of transmission vector specificities, increase in virulence and pathogenesis, the modification of tissue tropisms, the host invasion of first immunity and evolution of resistance to antivirals (Chin *et al.*, 2005; Shahzan *et al.*, 2019).

### Changes in virus due to recombination

Infection genomes can advance by recombination. Self recombination happens when two viral genomes recombine by homologous traverse. This occurs for DNA infections and is regular in prokaryotic infections, yet additionally for RNA infections or retroviruses (Murtaugh, Yuan and Faaberg, 2001). These recombination occasions can be of transformative preferred position for the infection when it assists with dodging host safe protections, for instance by changing surface protein antigenicity. Recombination with another living being happens when a viral genome recombines to gain arrangements from another living being. Normal determination can hold the obtained arrangement on the off chance that it gives a transformative preferred position to the infection and changes can adjust its unique capacities (Website, no date g). This occasion is regular in enormous dsDNA infections, some eukaryotic infections even gained numerous qualities from microscopic organisms. The outcomes of viral assessment Viral quasispecies: \*The quick and adaptable development of RNA infection genomes makes populace of infections which are on the whole extraordinary, called Viral quasispecies because a solitary succession can't depict precisely the viral populace in a solitary host or even in cell culture. \*Defective meddling virus: \*Defective meddling infections (DIs) emerge through erasure, adjustment, or recombination of a skilled viral genome. DIs are identified with satellite infection, yet not at all like them they are not monitored by normal determination, they show up and vanish (Girija *et al.*, 2019). DIs rival the viral genome for replication or potentially encapsidation factors and accordingly will, in general, weaken the infection and trigger host antiviral barriers. *Streptococcus mutans* is a potent initiator which causes dental caries. Essential oils are volatile compounds of plant secondary metabolism and act as a phytoprotective agent. Orange peel oil has an antibacterial activity on *Streptococcus mutans* (Website, no date h). These blemished genomes are amassing in cell culture, where the inborn antiviral guard is frequently damaged, in this way evacuating the negative determination of DIs. For instance, Sendai infection is utilized in cell culture to incite interferon, even though wild sort Sendai is closing off IFN creation. In reality, Sendai developed in cell culture is loaded with DIs, which are exceptionally strong to instigate inborn insusceptibility (Töpfer *et al.*, 2013). Our institution is passionate about high quality evidence based research and has excelled in various fields ( (Pc, Marimuthu and Devadoss, 2018; Ramesh *et al.*, 2018; Vijayashree Priyadharsini, Smiline Girija and Paramasivam, 2018; Ezhilarasan, Apoorva and Ashok Vardhan, 2019; Ramadurai *et al.*, 2019; Sridharan *et al.*, 2019; Vijayashree Priyadharsini, 2019; Chandrasekar *et al.*, 2020; Mathew *et al.*, 2020; R *et al.*, 2020; Samuel, 2021)

### CONCLUSION

Viral mutation rates are determined by multiple processes, including polymerase Intrinsic fidelity, triplication mode, endonuclease activity, spontaneous nucleic acid damage, access to post replicative repair, editing by host-encoded histamines, imbalance in force, template sequence content and template structure. Radiation of cell phone towers, fertilisers, food preservatives also causes a mutation in viruses. Further research can be done for finding the other factors that cause mutation in viruses. Mutation of viruses can have serious ill effects. We can prevent viruses by antiviral drugs and the primary method is by vaccination, which is intended to prevent outbreaks by building immunity to a virus or virus family. Hepatitis B is an infectious disease caused by Hepatitis B virus which affects the liver. Dental practitioners can be affected easily through several ways, by direct and indirect contact with blood, oral fluids, aerosols etc. Vaccination must be taken against the virus (Website, no date i). Vaccines are prepared using live viruses, killed viruses, or molecular subunits of the virus. The killed viral vaccines and subunit viruses are both incapable of causing disease (Marickar, Geetha and Neelakantan, 2014)

**REFERENCES**

1. Adelman, J. P. et al. (1987) 'Two mammalian genes transcribed from opposite strands of the same DNA locus', *Science*, 235(4795), pp. 1514–1517.
2. Anderson, J. P., Daifuku, R. and Loeb, L. A. (2004) 'Viral Error Catastrophe by Mutagenic Nucleosides', *Annual Review of Microbiology*, pp. 183–205. doi: 10.1146/annurev.micro.58.030603.123649.
3. Arenas, M. (2012) 'Simulation of molecular data under diverse evolutionary scenarios', *PLoS computational biology*, 8(5), p. e1002495.
4. Arenas, M. and Posada, D. (2010) 'The Effect of Recombination on the Reconstruction of Ancestral Sequences', *Genetics*, pp. 1133–1139. doi: 10.1534/genetics.109.113423.
5. Ashwin, K. S. and Muralidharan, N. P. (2015) 'Vancomycin-resistant enterococcus (VRE) vs Methicillin-resistant Staphylococcus Aureus (MRSA)', *Indian journal of medical microbiology*, 33 Suppl, pp. 166–167.
6. Awadalla, P. (2003) 'The evolutionary genomics of pathogen recombination', *Nature Reviews Genetics*, pp. 50–60. doi: 10.1038/nrg964.
7. Baltimore, D. (1971) 'Expression of animal virus genomes', *Bacteriological Reviews*, pp. 235–241. doi: 10.1128/mmbr.35.3.235-241.1971.
8. Barcellos-Hoff, M. H. and Brooks, A. L. (2001) 'Extracellular signaling through the microenvironment: a hypothesis relating carcinogenesis, bystander effects, and genomic instability', *Radiation research*, 156(5 Pt 2), pp. 618–627.
9. Bauch, C. T. et al. (2005) 'Dynamically Modeling SARS and Other Newly Emerging Respiratory Illnesses', *Epidemiology*, pp. 791–801. doi: 10.1097/01.ede.0000181633.80269.4c.
10. Belshaw, R., Sanjuán, R. and Pybus, O. G. (2011) 'Viral mutation and substitution: units and levels', *Current Opinion in Virology*, pp. 430–435. doi: 10.1016/j.coviro.2011.08.004.
11. Biswas, S. J. and Khuda-Bukhsh, A. R. (2005) 'Cytotoxic and genotoxic effects of the azo-dye p-dimethylaminoazobenzene in mice: A time-course study', *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, pp. 1–8. doi: 10.1016/j.mrgentox.2005.06.011.
12. Bootsma, M. C. J. (2007) The Effect of Public Health Measures on the 1918 Influenza Pandemic in U.S. Cities.
13. Bull, J. J., Sanjuán, R. and Wilke, C. O. (2007) 'Theory of Lethal Mutagenesis for Viruses', *Journal of Virology*, pp. 2930–2939. doi: 10.1128/jvi.01624-06.
14. Cao, Y. et al. (2016) 'Resistance-associated mutations to HCV protease inhibitors naturally pre-existed in HIV/HCV coinfecting, treatment-naïve patients', *Clinics and Research in Hepatology and Gastroenterology*, pp. 597–604. doi: 10.1016/j.clinre.2016.02.004.
15. Chandrasekar, R. et al. (2020) 'Development and validation of a formula for objective assessment of cervical vertebral bone age', *Progress in orthodontics*, 21(1), p. 38.
16. Chang, W. P. and Little, J. B. (1991) 'Delayed reproductive death in X-irradiated Chinese hamster ovary cells', *International journal of radiation biology*, 60(3), pp. 483–496.
17. Chin, M. P. S. et al. (2005) 'Identification of a major restriction in HIV-1 intersubtype recombination', *Proceedings of the National Academy of Sciences*, pp. 9002–9007. doi: 10.1073/pnas.0502522102.
18. Clutton, S. M. et al. (1996) 'Radiation-induced genomic instability and persisting oxidative stress in primary bone marrow cultures', *Carcinogenesis*, 17(8), pp. 1633–1639.
19. Cremer, M. et al. (2001) 'Non-random radial higher-order chromatin arrangements in nuclei of diploid human cells', *Chromosome research: an international journal on the molecular, supramolecular and evolutionary aspects of chromosome biology*, 9(7), pp. 541–567.
20. Crotty, S., Cameron, C. E. and Andino, R. (2001) 'RNA virus error catastrophe: Direct molecular test by using ribavirin', *Proceedings of the National Academy of Sciences*, pp. 6895–6900. doi: 10.1073/pnas.111085598.
21. Deogade, S., Gupta, P. and Ariga, P. (2018) 'Effect of monopoly-coating agent on the surface roughness of a tissue conditioner subjected to cleansing and disinfection: A Contact Profilometric In vitro study', *Contemporary Clinical Dentistry*, p. 122. doi: 10.4103/ccd.ccd\_112\_18.
22. Domingo, E. et al. (2006) 'Viruses as Quasispecies: Biological Implications', *Current Topics in Microbiology and Immunology*, pp. 51–82. doi: 10.1007/3-540-26397-7\_3.
23. Dua, K. et al. (2019) 'The potential of siRNA based drug delivery in respiratory disorders: Recent advances and progress', *Drug development research*, 80(6), pp. 714–730.
24. Duffy, S., Shackelton, L. A. and Holmes, E. C. (2008) 'Rates of evolutionary change in viruses: patterns and determinants', *Nature Reviews Genetics*, pp. 267–276. doi: 10.1038/nrg2323.
25. Duraisamy, R. et al. (2019) 'Compatibility of Nonoriginal Abutments With Implants: Evaluation of Microgap at the Implant-Abutment Interface, With Original and Nonoriginal Abutments', *Implant dentistry*, 28(3), pp. 289–295.
26. Ezhilarasan, D. (2018) 'Oxidative stress is bane in chronic liver diseases: Clinical and experimental

- perspective', Arab journal of gastroenterology: the official publication of the Pan-Arab Association of Gastroenterology, 19(2), pp. 56–64.
27. Ezhilarasan, D., Apoorva, V. S. and Ashok Vardhan, N. (2019) 'Syzygium cumini extract induced reactive oxygen species-mediated apoptosis in human oral squamous carcinoma cells', Journal of oral pathology & medicine: official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology, 48(2), pp. 115–121.
  28. Ezhilarasan, D., Sokal, E. and Najimi, M. (2018) 'Hepatic fibrosis: It is time to go with hepatic stellate cell-specific therapeutic targets', Hepatobiliary & pancreatic diseases international: HBPD INT, 17(3), pp. 192–197.
  29. Fadel, H. J. and Poeschla, E. M. (2011) 'Retroviral restriction and dependency factors in primates and carnivores', Veterinary Immunology and Immunopathology, pp. 179–189. doi: 10.1016/j.vetimm.2011.06.002.
  30. Ferguson, N. M. et al. (2005) 'Strategies for containing an emerging influenza pandemic in Southeast Asia', Nature, pp. 209–214. doi: 10.1038/nature04017.
  31. Fouchier, R. et al. (2005) 'Global task force for influenza', Nature, pp. 419–420. doi: 10.1038/435419a.
  32. Fraser, C. et al. (2004) 'Factors that make an infectious disease outbreak controllable', Proceedings of the National Academy of Sciences, pp. 6146–6151. doi: 10.1073/pnas.0307506101.
  33. Garcin, D., Itoh, M. and Kolakofsky, D. (1997) 'A Point Mutation in the Sendai Virus Accessory C Proteins Attenuates Virulence for Mice, but Not Virus Growth in Cell Culture', Virology, pp. 424–431. doi: 10.1006/viro.1997.8836.
  34. Geetha, R. V., Thangavelu, L. and Others (2019) 'Evaluation of anti-inflammatory action of Laurus nobilis-an in vitro study', International Journal of Research in Pharmaceutical Sciences, 10(2), pp. 1209–1213.
  35. Gheena, S. and Ezhilarasan, D. (2019) 'Syringic acid triggers reactive oxygen species-mediated cytotoxicity in HepG2 cells', Human & experimental toxicology, 38(6), pp. 694–702.
  36. Gilbert, C. et al. (2016) 'Continuous Influx of Genetic Material from Host to Virus Populations', PLOS Genetics, p. e1005838. doi: 10.1371/journal.pgen.1005838.
  37. Giriya, A. S. S. et al. (2019) 'Plasmid-encoded resistance to trimethoprim/sulfamethoxazole mediated by dfrA1, dfrA5, sul1 and sul2 among Acinetobacter baumannii isolated from urine samples of patients with severe urinary tract infection', Journal of Global Antimicrobial Resistance, pp. 145–146. doi: 10.1016/j.jgar.2019.04.001.
  38. Giriya, S. A., Priyadharsini, J. V. and Paramasivam, A. (2019) 'Prevalence of carbapenem-hydrolyzing OXA-type  $\beta$ -lactamases among Acinetobacter baumannii in patients with severe urinary tract infection', Acta Microbiologica et Immunologica Hungarica, pp. 1–7. doi: 10.1556/030.66.2019.030.
  39. Gomathi, A. C. et al. (2020) 'Anticancer activity of silver nanoparticles synthesized using aqueous fruit shell extract of Tamarindus indica on MCF-7 human breast cancer cell line', Journal of Drug Delivery Science and Technology, p. 101376. doi: 10.1016/j.jddst.2019.101376.
  40. Graci, J. D. et al. (2007) 'Lethal mutagenesis of poliovirus mediated by a mutagenic pyrimidine analogue', Journal of virology, 81(20), pp. 11256–11266.
  41. Hatchett, R. J., Mecher, C. E. and Lipsitch, M. (2007) 'Public health interventions and epidemic intensity during the 1918 influenza pandemic', Proceedings of the National Academy of Sciences of the United States of America, 104(18), pp. 7582–7587.
  42. Hill, M. A. (1999) 'Radiation damage to DNA: the importance of track structure', Radiation measurements, 31(1-6), pp. 15–23.
  43. Holland, J. J. et al. (1990) 'Mutation Frequencies at Defined Single Codon Sites in Vesicular Stomatitis Virus and Poliovirus Can Be Increased Only Slightly by Chemical Mutagenesis', Journal of Virology, pp. 3960–3962. doi: 10.1128/jvi.64.8.3960-3962.1990.
  44. Hughes, A. L. and Friedman, R. (2005) 'Poxvirus genome evolution by gene gain and loss', Molecular Phylogenetics and Evolution, pp. 186–195. doi: 10.1016/j.ympev.2004.12.008.
  45. Jeevanandan, G. and Govindaraju, L. (2018) 'Clinical comparison of Kedo-S paediatric rotary files vs manual instrumentation for root canal preparation in primary molars: a double blinded randomised clinical trial', European Archives of Paediatric Dentistry, pp. 273–278. doi: 10.1007/s40368-018-0356-6.
  46. Jefferson, T. et al. (2009) 'Physical interventions to interrupt or reduce the spread of respiratory viruses: systematic review', BMJ, pp. b3675–b3675. doi: 10.1136/bmj.b3675.
  47. J, P. C. et al. (2018) 'Prevalence and measurement of anterior loop of the mandibular canal using CBCT: A cross sectional study', Clinical implant dentistry and related research, 20(4), pp. 531–534.
  48. Khanna, K. K. et al. (2001) 'ATM, a central controller of cellular responses to DNA damage', Cell Death & Differentiation, pp. 1052–1065. doi: 10.1038/sj.cdd.4400874.
  49. Liang, L. et al. (2002) 'Radiation-induced genetic instability in vivo depends on p53 status', Mutation research, 502(1-2), pp. 69–80.

50. Malim, M. H. and Bieniasz, P. D. (2012) 'HIV Restriction Factors and Mechanisms of Evasion', *Cold Spring Harbor perspectives in medicine*, 2(5), p. a006940.
51. Malli Sureshbabu, N. et al. (2019) 'Concentrated Growth Factors as an Ingenious Biomaterial in Regeneration of Bony Defects after Periapical Surgery: A Report of Two Cases', *Case reports in dentistry*, 2019, p. 7046203.
52. Marickar, R. F., Geetha, R. V. and Neelakantan, P. (2014) 'Efficacy of Contemporary and Novel Intracanal Medicaments against *Enterococcus Faecalis*', *Journal of Clinical Pediatric Dentistry*, pp. 47–50. doi: 10.17796/jcpd.39.1.wmw9768314h56666.
53. Martina, B. E. E. et al. (2003) 'SARS virus infection of cats and ferrets', *Nature*, pp. 915–915. doi: 10.1038/425915a.
54. Mathew, M. G. et al. (2020) 'Evaluation of adhesion of *Streptococcus mutans*, plaque accumulation on zirconia and stainless steel crowns, and surrounding gingival inflammation in primary molars: Randomized controlled trial', *Clinical oral investigations*, pp. 1–6.
55. Mehta, M. et al. (2019) 'Oligonucleotide therapy: An emerging focus area for drug delivery in chronic inflammatory respiratory diseases', *Chemico-biological interactions*, 308, pp. 206–215.
56. Menon, S. et al. (2018) 'Selenium nanoparticles: A potent chemotherapeutic agent and an elucidation of its mechanism', *Colloids and Surfaces B: Biointerfaces*, pp. 280–292. doi: 10.1016/j.colsurfb.2018.06.006.
57. Meyerson, N. R. and Sawyer, S. L. (2011) 'Two-stepping through time: mammals and viruses', *Trends in Microbiology*, pp. 286–294. doi: 10.1016/j.tim.2011.03.006.
58. Murtaugh, M. P., Yuan, S. and Faaberg, K. S. (2001) 'Appearance of Novel PRRSV Isolates by Recombination in the Natural Environment', *Advances in Experimental Medicine and Biology*, pp. 31–36. doi: 10.1007/978-1-4615-1325-4\_4.
59. Panchal, V., Jeevanandan, G. and Subramanian, E. M. G. (2019) 'Comparison of post-operative pain after root canal instrumentation with hand K-files, H-files and rotary Kedo-S files in primary teeth: a randomised clinical trial', *European archives of paediatric dentistry: official journal of the European Academy of Paediatric Dentistry*, 20(5), pp. 467–472.
60. Pandir, D. (2016) 'DNA damage in human germ cell exposed to the some food additives in vitro', *Cytotechnology*, pp. 725–733. doi: 10.1007/s10616-014-9824-y.
61. Paramasivam, A., Vijayashree Priyadharsini, J. and Raghunandhakumar, S. (2020) 'N6-adenosine methylation (m6A): a promising new molecular target in hypertension and cardiovascular diseases', *Hypertension research: official journal of the Japanese Society of Hypertension*, 43(2), pp. 153–154.
62. Pc, J., Marimuthu, T. and Devadoss, P. (2018) 'Prevalence and measurement of anterior loop of the mandibular canal using CBCT: A cross sectional study', *Clinical implant dentistry and related research*. Available at: <https://europepmc.org/article/med/29624863>.
63. Peace-Brewer, A. L. et al. (1996) 'A Point Mutation in HLA-A\*0201 Results in Failure to Bind the TAP Complex and to Present Virus-Derived Peptides to CTL', *Immunity*, pp. 505–514. doi: 10.1016/s1074-7613(00)80416-1.
64. Perelson, A. S. (2018) *Theoretical Immunology, Part Two*. CRC Press.
65. Planelles, V. (2012) 'SAMHD1 Joins the Red Queen's Court', *Cell Host & Microbe*, pp. 103–105. doi: 10.1016/j.chom.2012.02.001.
66. Podolin, P. L. et al. (2002) 'A Potent and Selective Nonpeptide Antagonist of CXCR2 Inhibits Acute and Chronic Models of Arthritis in the Rabbit', *The Journal of Immunology*, pp. 6435–6444. doi: 10.4049/jimmunol.169.11.6435.
67. Prabakar, J. et al. (2018) 'Comparative Evaluation of Retention, Cariostatic Effect and Discoloration of Conventional and Hydrophilic Sealants - A Single Blinded Randomized Split Mouth Clinical Trial', *Contemporary clinical dentistry*, 9(Suppl 2), pp. S233–S239.
68. Priyadharsini, J. V. et al. (2018) 'An insight into the emergence of *Acinetobacter baumannii* as an orodental pathogen and its drug resistance gene profile – An in silico approach', *Heliyon*, p. e01051. doi: 10.1016/j.heliyon.2018.e01051.
69. Rajendran, R. et al. (2019) 'Comparative Evaluation of Remineralizing Potential of a Paste Containing Bioactive Glass and a Topical Cream Containing Casein Phosphopeptide-Amorphous Calcium Phosphate: An in Vitro Study', *Pesquisa Brasileira em Odontopediatria e Clínica Integrada*, pp. 1–10. doi: 10.4034/pboci.2019.191.61.
70. Rajeshkumar, S. et al. (2018) 'Biosynthesis of zinc oxide nanoparticles using *Mangifera indica* leaves and evaluation of their antioxidant and cytotoxic properties in lung cancer (A549) cells', *Enzyme and microbial technology*, 117, pp. 91–95.
71. Rajeshkumar, S. et al. (2019) 'Antibacterial and antioxidant potential of biosynthesized copper nanoparticles mediated through *Cissampelos grandifolia* plant extract', *Journal of photochemistry and photobiology. B, Biology*, 197, p. 111531.
72. Ramadurai, N. et al. (2019) 'Effectiveness of 2% Articaine as an anesthetic agent in children: randomized



- controlled trial', *Clinical oral investigations*, 23(9), pp. 3543–3550.
73. Ramakrishnan, M., Dhanalakshmi, R. and Subramanian, E. M. G. (2019) 'Survival rate of different fixed posterior space maintainers used in Paediatric Dentistry - A systematic review', *The Saudi dental journal*, 31(2), pp. 165–172.
  74. Ramesh, A. et al. (2018) 'Comparative estimation of sulfiredoxin levels between chronic periodontitis and healthy patients - A case-control study', *Journal of periodontology*, 89(10), pp. 1241–1248.
  75. R, H. et al. (2020) 'CYP2 C9 polymorphism among patients with oral squamous cell carcinoma and its role in altering the metabolism of benzo[a]pyrene', *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*, pp. 306–312. doi: 10.1016/j.oooo.2020.06.021.
  76. Samuel, S. R. (2021) 'Can 5-year-olds sensibly self-report the impact of developmental enamel defects on their quality of life?', *International journal of paediatric dentistry / the British Paedodontic Society [and] the International Association of Dentistry for Children*, 31(2), pp. 285–286.
  77. Samuel, S. R., Acharya, S. and Rao, J. C. (2020) 'School Interventions-based Prevention of Early-Childhood Caries among 3-5-year-old children from very low socioeconomic status: Two-year randomized trial', *Journal of public health dentistry*, 80(1), pp. 51–60.
  78. Sanjuán, R. (2016) 'Viral Mutation Rates', *Virus Evolution: Current Research and Future Directions*, pp. 1–28. doi: 10.21775/9781910190234.01.
  79. Sayah, D. M. et al. (2004) 'Cyclophilin A retrotransposition into TRIM5 explains owl monkey resistance to HIV-1', *Nature*, pp. 569–573. doi: 10.1038/nature02777.
  80. Schotsaert, M. and García-Sastre, A. (2014) 'Influenza Vaccines: A Moving Interdisciplinary Field', *Viruses*, pp. 3809–3826. doi: 10.3390/v6103809.
  81. Shahzan, M. S. et al. (2019) 'A computational study targeting the mutated L321F of ERG11 gene in *C. albicans*, associated with fluconazole resistance with bioactive compounds from *Acacia nilotica*', *Journal de Mycologie Médicale*, pp. 303–309. doi: 10.1016/j.mycmed.2019.100899.
  82. Sharma, P. et al. (2019) 'Emerging trends in the novel drug delivery approaches for the treatment of lung cancer', *Chemico-biological interactions*, 309, p. 108720.
  83. Sridharan, G. et al. (2019) 'Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma', *Journal of oral pathology & medicine: official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*, 48(4), pp. 299–306.
  84. Stremlau, M. et al. (2005) 'Species-Specific Variation in the B30.2(SPRY) Domain of TRIM5 $\alpha$  Determines the Potency of Human Immunodeficiency Virus Restriction', *Journal of Virology*, pp. 3139–3145. doi: 10.1128/jvi.79.5.3139-3145.2005.
  85. Tanaka, R. (2007) 'INHIBITORY EFFECTS OF XANTHONE ON PARAQUAT- AND NaNO<sub>2</sub>-INDUCED GENOTOXICITY IN CULTURED CELLS', *The Journal of Toxicological Sciences*, pp. 571–574. doi: 10.2131/jts.32.571.
  86. Töpfer, A. et al. (2013) 'Probabilistic Inference of Viral Quasispecies Subject to Recombination', *Journal of Computational Biology*, pp. 113–123. doi: 10.1089/cmb.2012.0232.
  87. Varghese, S. S., Ramesh, A. and Veeraiyan, D. N. (2019) 'Blended Module-Based Teaching in Biostatistics and Research Methodology: A Retrospective Study with Postgraduate Dental Students', *Journal of dental education*, 83(4), pp. 445–450.
  88. Vijayashree Priyadharsini, J. (2019) 'In silico validation of the non-antibiotic drugs acetaminophen and ibuprofen as antibacterial agents against red complex pathogens', *Journal of periodontology*, 90(12), pp. 1441–1448.
  89. Vijayashree Priyadharsini, J., Smiline Girija, A. S. and Paramasivam, A. (2018) 'In silico analysis of virulence genes in an emerging dental pathogen *A. baumannii* and related species', *Archives of oral biology*, 94, pp. 93–98.
  90. Vishnu Prasad, S. et al. (2018) 'Report on oral health status and treatment needs of 5-15 years old children with sensory deficits in Chennai, India', *Special care in dentistry: official publication of the American Association of Hospital Dentists, the Academy of Dentistry for the Handicapped, and the American Society for Geriatric Dentistry*, 38(1), pp. 58–59.
  91. Wahab, P. U. A. et al. (2018) 'Scalpel Versus Diathermy in Wound Healing After Mucosal Incisions: A Split-Mouth Study', *Journal of oral and maxillofacial surgery: official journal of the American Association of Oral and Maxillofacial Surgeons*, 76(6), pp. 1160–1164.
  92. Website (no date a). Available at: Shahana RY, Muralidharan NP. Efficacy of mouth rinse in maintaining oral health of patients attending orthodontic clinics. *Research Journal of Pharmacy and Technology* 2016;9:1991. <https://doi.org/10.5958/0974-360x.2016.00406.6>. (Accessed: 16 June 2020).
  93. Website (no date b). Available at: Selvakumar R, Np M. COMPARISON IN BENEFITS OF HERBAL MOUTHWASHES WITH CHLORHEXIDINE MOUTHWASH: A REVIEW. *Asian Journal of Pharmaceutical and Clinical Research* 2017;10:3. <https://doi.org/10.22159/ajpcr.2017.v10i2.13304>. (Accessed: 16 June 2020).

94. Website (no date c). Available at: Smiline ASG, Vijayashree JP, Paramasivam A. Molecular characterization of plasmid-encoded blaTEM, blaSHV and blaCTX-M among extended-spectrum against enterococcus] producing Acinetobacter baumannii. *British Journal of Biomedical Science* 2018;75:200–2. <https://doi.org/10.1080/09674845.2018.1492207>. (Accessed: 16 June 2020).
95. Website (no date d). Available at: Girija SAS, Jayaseelan VP, Arumugam P. Prevalence of VIM- and GIM-producing Acinetobacter baumannii from patients with severe urinary tract infection. *Acta Microbiologica et Immunologica Hungarica* 2018;65:539–50. <https://doi.org/10.1556/030.65.2018.038>. (Accessed: 16 June 2020).
96. Website (no date e). Available at: Priyadharsini JV, Vijayashree Priyadharsini J, Smiline Girija AS, Paramasivam A. In silico analysis of virulence genes in an emerging dental pathogen A. baumannii and related species. *Archives of Oral Biology* 2018;94:93–8. <https://doi.org/10.1016/j.archoralbio.2018.07.001>. (Accessed: 16 June 2020).
97. Website (no date f). Available at: Holzerlandt, R., C. Orengo, P. Kellam, and M. M. Alba. 2002. 'Identification of New Herpesvirus Gene Homologs in the Human Genome.' *Genome Research*. <http://genome.cshlp.org/content/12/11/1739.short>. (Accessed: 8 June 2020).
98. Website (no date g). Available at: Neher, Richard A., and Thomas Leitner. 2010. 'Recombination Rate and Selection Strength in HIV Intra-Patient Evolution.' *PLoS Computational Biology* 6 (1). <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2813257/>. (Accessed: 8 June 2020).
99. Website (no date h). Available at: Vaishali M, Geetha RV. Antibacterial activity of Orange peel oil on Streptococcus mutans and Enterococcus-An In-vitro study. *Research Journal of Pharmacy and Technology* 2018;11:513. <https://doi.org/10.5958/0974-360x.2018.00094.x>. (Accessed: 16 June 2020).
100. Website (no date i). Available at: Pratha AA, Ashwatha Pratha A, Geetha RV. Awareness on Hepatitis-B vaccination among dental students-A Questionnaire Survey. *Research Journal of Pharmacy and Technology* 2017;10:1360. <https://doi.org/10.5958/0974-360x.2017.00240.2>. (Accessed: 16 June 2020).
101. Yılmaz, S. et al. (2008) 'Clastogenic effects of food additive citric acid in human peripheral lymphocytes', *Cytotechnology*, pp. 137–144. doi: 10.1007/s10616-008-9137-0.
102. Yılmaz, S., Ünal, F. and Yüzbaşıoğlu, D. (2009) 'The in vitro genotoxicity of benzoic acid in human peripheral blood lymphocytes', *Cytotechnology*, 60(1-3), p. 55.
103. Zengin, N. et al. (2011) 'The evaluation of the genotoxicity of two food preservatives: Sodium benzoate and potassium benzoate', *Food and Chemical Toxicology*, pp. 763–769. doi: 10.1016/j.fct.2010.11.040.